METHOD



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Derivation of HLA types from shotgun sequence datasets

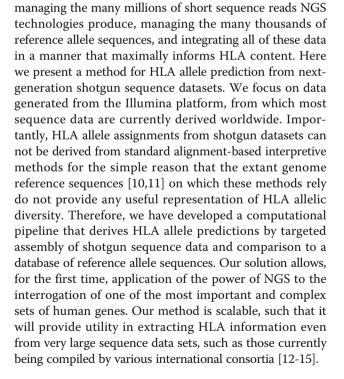
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Abstract

The human leukocyte antigen (HLA) is key to many aspects of human physiology and medicine. All current sequencebased HLA typing methodologies are targeted approaches requiring the amplification of specific HLA gene segments. Whole genome, exome and transcriptome shotgun sequencing can generate prodigious data but due to the complexity of HLA loci these data have not been immediately informative regarding HLA genotype. We describe HLAminer, a computational method for identifying HLA alleles directly from shotgun sequence datasets (http://www. bcgsc.ca/platform/bioinfo/software/hlaminer). This approach circumvents the additional time and cost of generating HLA-specific data and capitalizes on the increasing accessibility and affordability of massively parallel sequencing.

Background

Due to its central role in adaptive immunity, human leukocyte antigen (HLA) is implicated in wide ranging areas of medicine, from infectious disease and vaccinology to cancer, autoimmunity, aging and regenerative and transplantation medicine [1-7]. The HLA locus is the most polymorphic region of the genome with over 5,000 variant HLA-class I allelic sequences catalogued to date. This genetic heterogeneity is the principal challenge to HLA typing methodologies, and it is the reason why this region has remained largely opaque to analysis by next-generation sequencing (NGS) platforms. Conventional sequence-based HLA typing approaches, the most recent of which exploits the sequence throughput of the Illumina MiSeq [8] and relatively long sequence reads of the 454 NGS platform [9], are targeted assays that rely on amplification of hypervariable sub-regions of these loci and variant detection within these amplicons. As such, HLA calls are based on sequence information that is not as comprehensive as for shotgun datasets, and must be generated *de novo* for each subject. The widespread uptake of large-scale genome, exome and transcriptome shotgun sequencing approaches for biomedical research, and now for clinical use, prompted us to explore the utility of these types of NGS data sets for HLA typing. The need has been for a solution to the problem of



Materials and methods

Library construction and sequencing

Written informed consent was obtained from all donors and samples were collected following assessment of tissue specimens by a pathologist according to standardized



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